

# BCHE2030 TUTORIAL 3

**Group**

# CONTENTS

Proteins:

amino acid characteristic

Pka value and ionization properties

Protein structure:

Amino acid sequence & Homologous protein, Protein Folding  
(Protein II p1-12, p27-35)

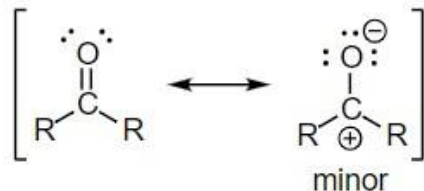
Secondary, tertiary structure

# 1. HOW CAN I DISTINGUISH WHETHER THE CARBON ATOM IN AN ALDEHYDE GROUP IS NEUTRAL, POSITIVELY OR NEGATIVELY CHARGED?

The carbonyl carbon can be neutral or carry a partial positive charge due to the presence of a *more* electronegative oxygen atom

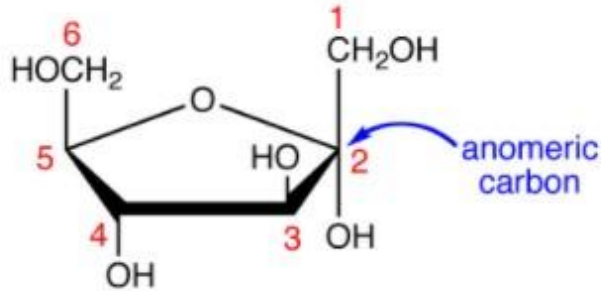


The carbonyl carbon in aldehyde has two resonance structures.

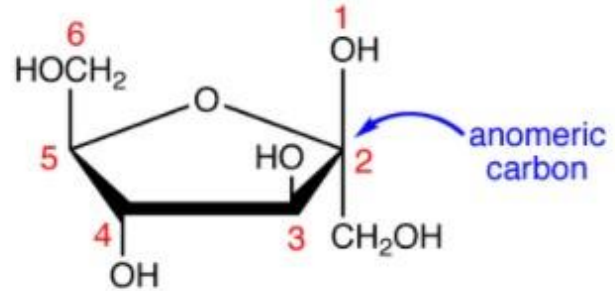


## 2. WHAT ARE ALPHA AND BETA ANOMERS?

In alpha anomers, the hydroxyl group bonded to the anomeric carbon is pointing downwards whereas in beta anomers, it is pointing upwards.



$\alpha$ -D-fructofuranose

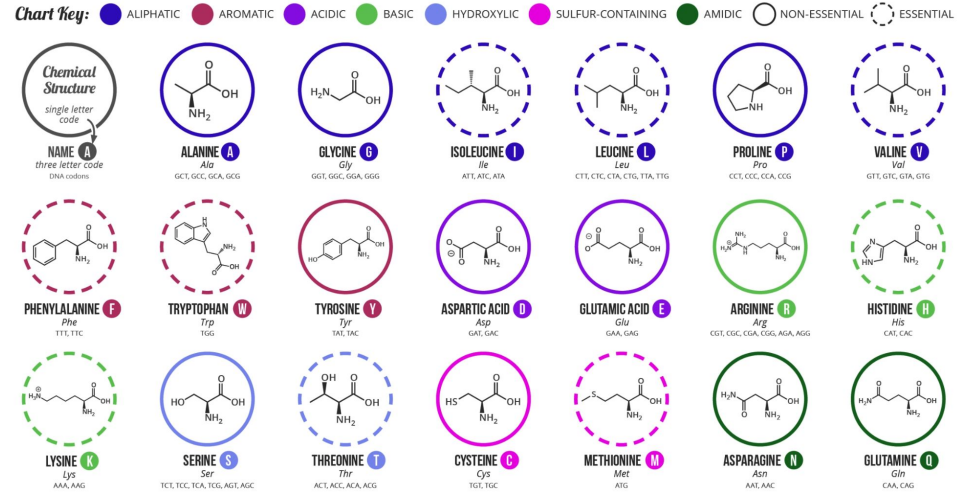


$\beta$ -D-fructofuranose

# 20 AMINO ACID

- Hydrophobic amino acids
- Polar amino acids
- -> +ve charged
- -> -ve charged

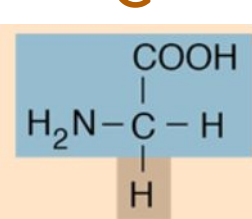
Amino Acids Are The Building Blocks Of Proteins In Living Organisms. There Are Over 500 Amino Acids Found In Nature - However, The Human Genetic Code Only Directly Encodes 20. 'essential' Amino Acids Must Be Obtained From The Diet, Whilst Non-essential Amino Acids Can Be Synthesised In The Body.



# HYDROPHOBIC AMINO ACID

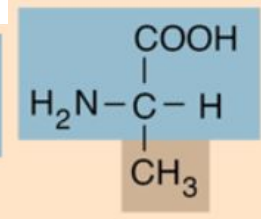
1. R group have similar electronegativity → hydrophobic
2. **L** & **I** have same chemical component
3. **P** aliphatic ring restrict in peptide angle
4. **M** is code for start codon (AUG)

**G**



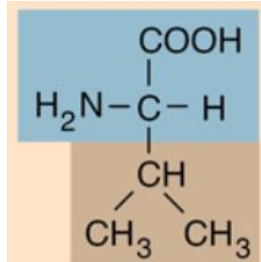
Glycine (gly)

**A**



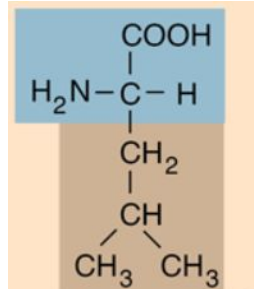
Alanine (ala)

**V**



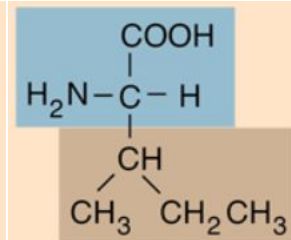
Valine (val)\*

**L**



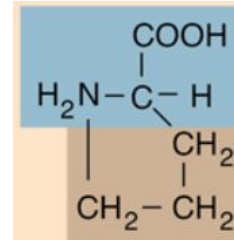
Leucine (leu)\*

**I**



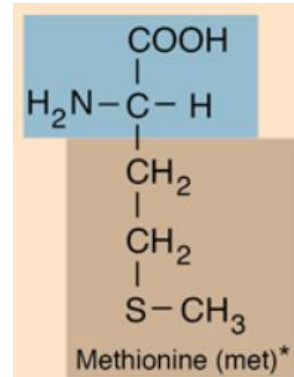
Isoleucine (ile)\*

**P**



Proline (pro)

**M**

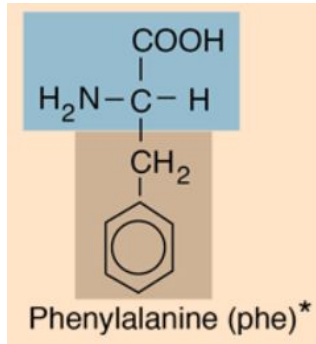


Methionine (met)\*

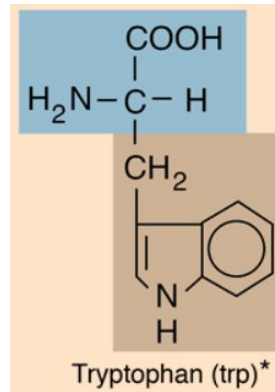
# AROMATIC AMINO ACID

aromatic ring on the side chain of each of these three amino acids

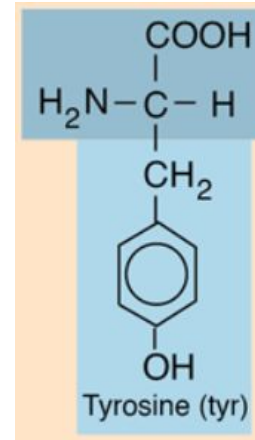
F



W



Y



---

**aromatic**

---

**hydrophobic**

---

**polar**

# POLAR (NERUTAL) AMINO ACID

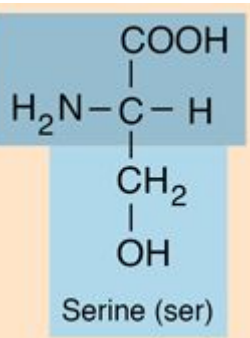
common phosphorylation site of protein

Nitrogen carrier

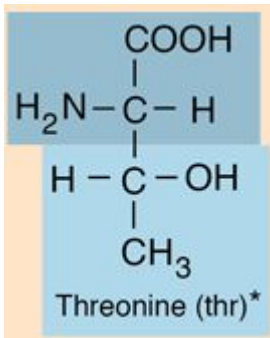
Disulfur bond formation

Intra-Inter bond

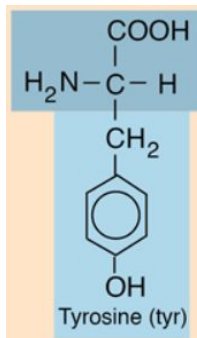
S



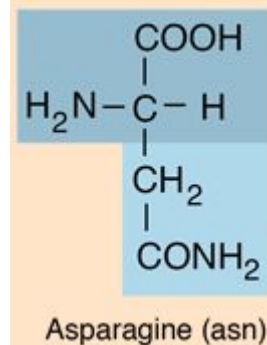
T



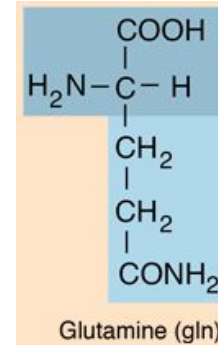
Y



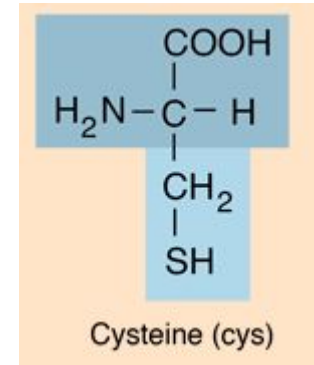
N



Q



C

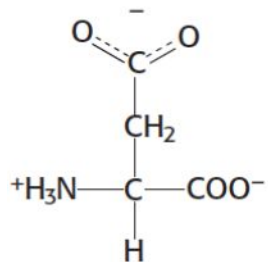




## POLAR (-VE CHARGED) AMINO ACID

$$pI < pH(7.4)$$

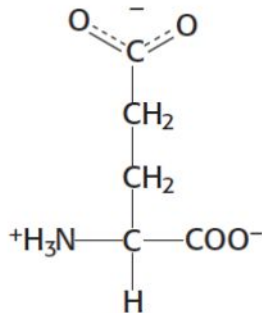
**D**



Aspartate  
(Asp, D)

3.7

**E**



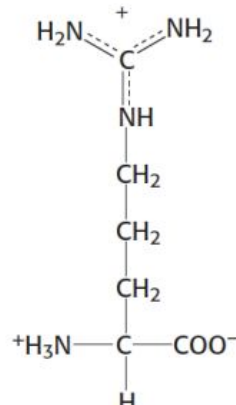
Glutamate  
(Glu, E)

4

## POLAR (+VE CHARGED) AMINO ACID

$$pI > pH(7.4)$$

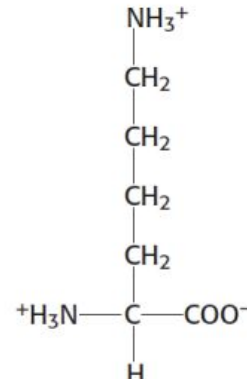
**R**



Arginine  
(Arg, R)

12.5

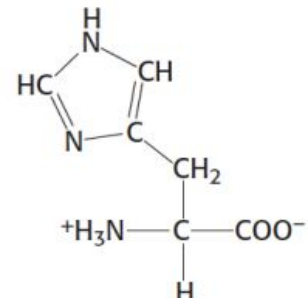
**K**



Lysine  
(Lys, K)

10.5

**H**



Histidine  
(His, H)

6

# PKA VALUE

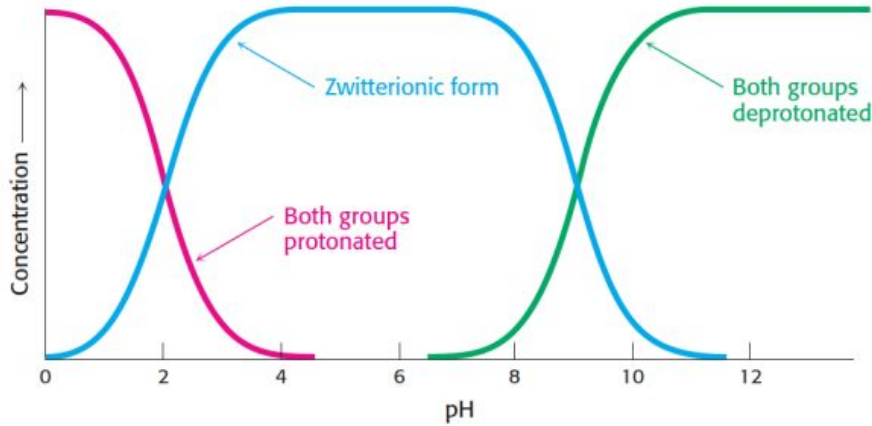
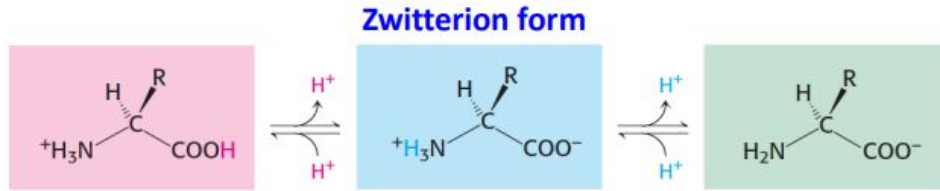
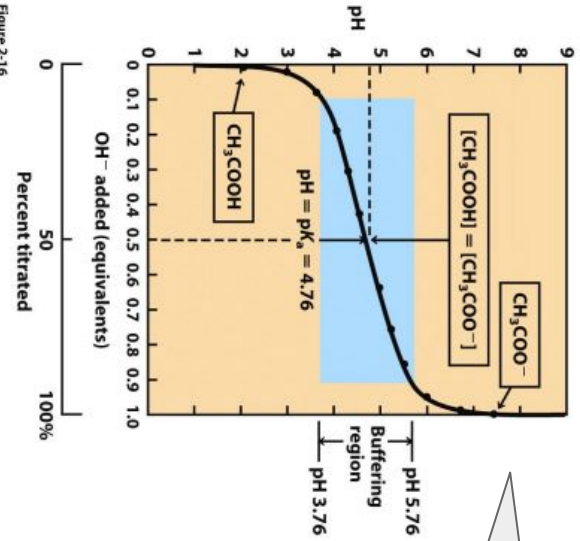


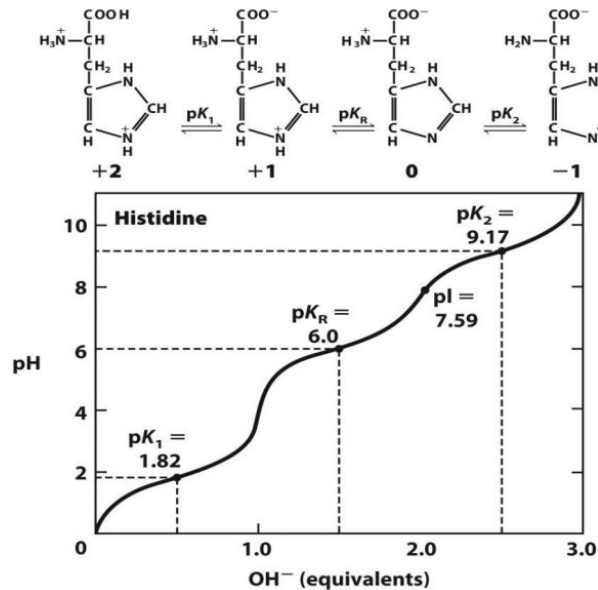
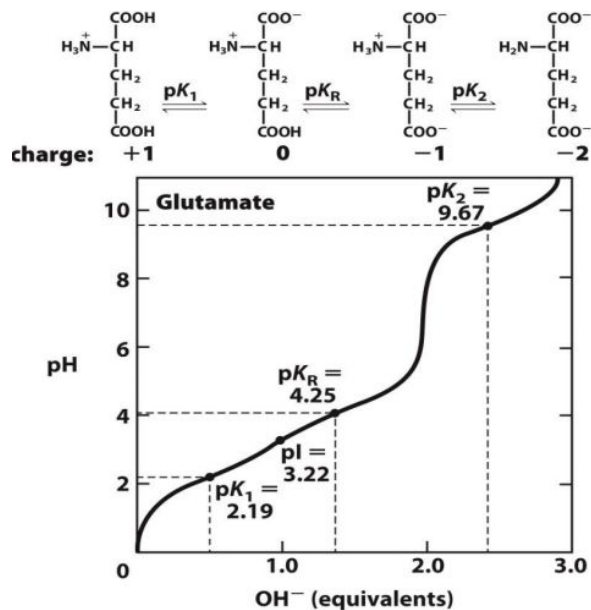
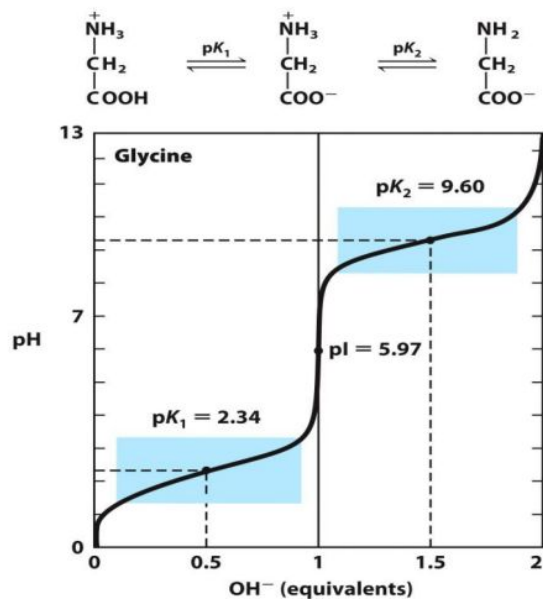
Figure of pH.  
altered form pr

Figure 2-16  
Lehninger Principles of Biochemistry, Fifth Edition  
© 2008 W. H. Freeman and Company



Recall  
Water  
Chapter

# PKA VALUE

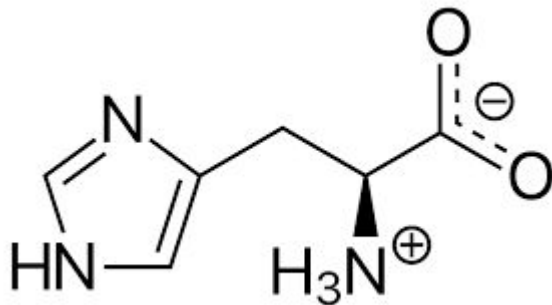
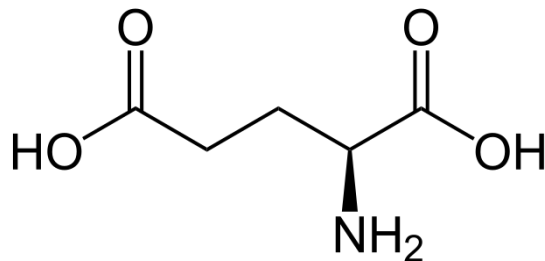
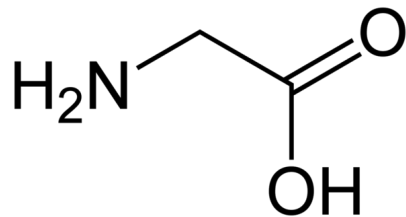


# ISOELECTRIC POINT

$$pK_1 + pK_2 / 2$$

$$pK_1 + pK_R / 2$$

$$pK_R + pK_2 / 2$$



# ISOLECTRIC POINT (DEFINITIONS)

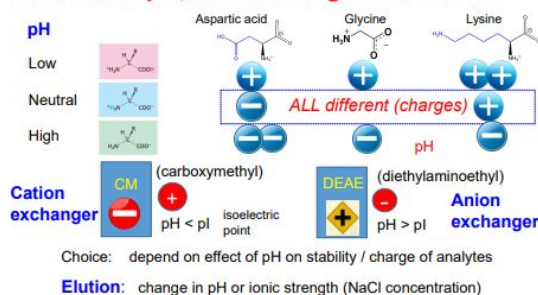
The characteristic pH at which the net charge of the molecule is zero.

Given by the average of the pKas that involve the zwitterion.

The pH at which the amino acid does NOT migrate in an electric field. (Laboratory applications)

Recall zwitterion: able to act as either an acid or a base (lecture slide 27)

## Choice of pH, Ion-exchanger and Eluent



source: BCHE3030

## Isoelectric Focusing (IEF)

### Simple electrophoresis

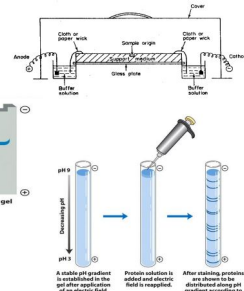
- separation by charge and mass
- different mobilities at a particular pH

### SDS-PAGE

- separation by **size only**

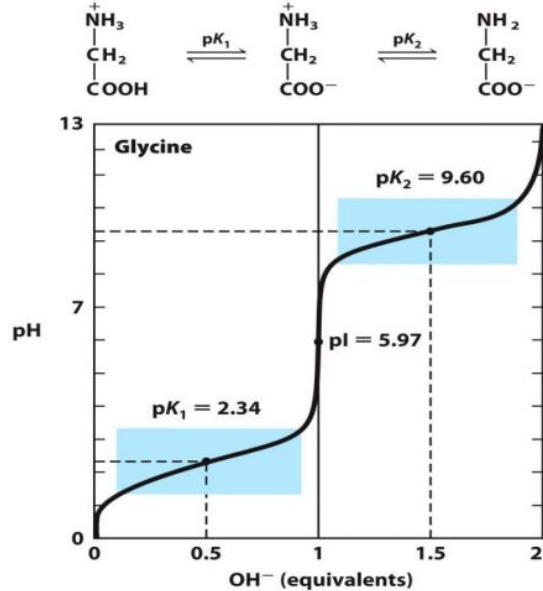
### Isoelectric focusing

- separation by **charge only**
- presence of pH gradient
- sample stops at the end
- steady state electrophoresis

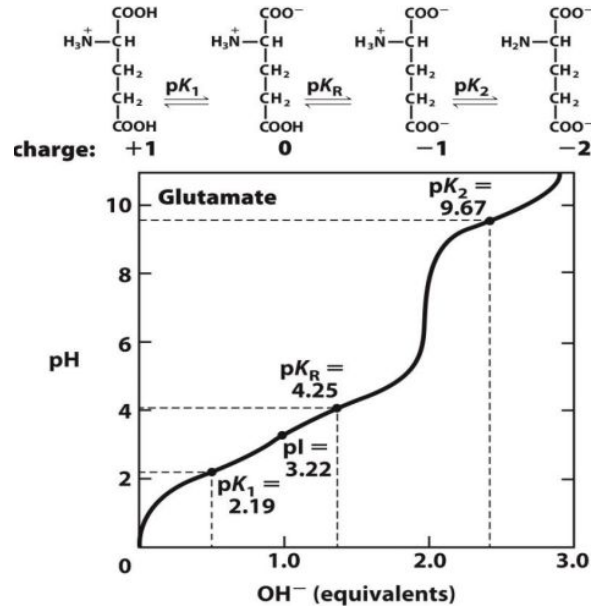


# ISOELECTRIC POINT

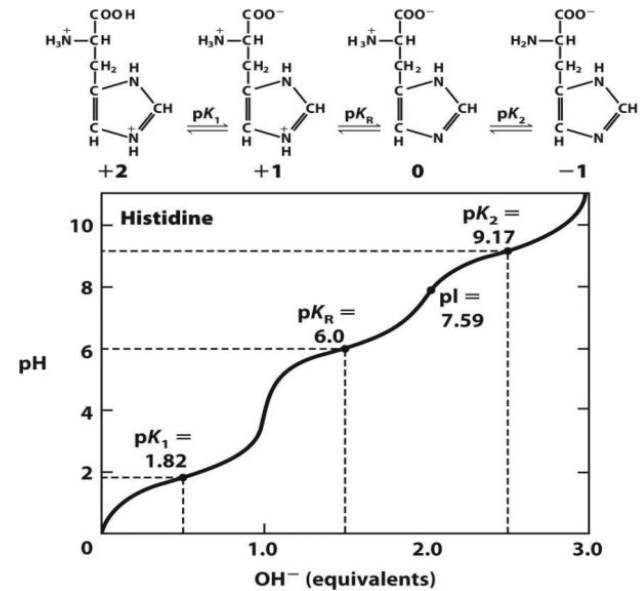
$$(pk_1 + pk_2) / 2$$



$$(pk_1 + pk_R) / 2$$

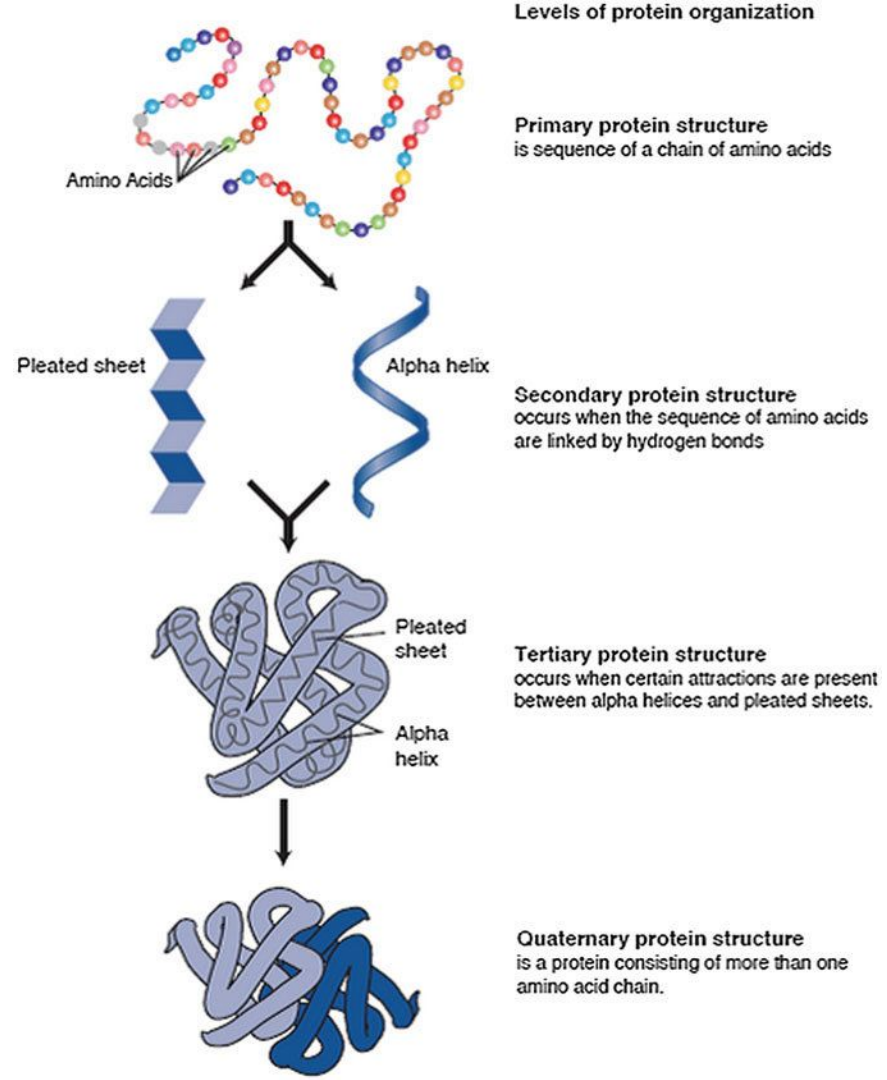


$$(pk_R + pk_2) / 2$$



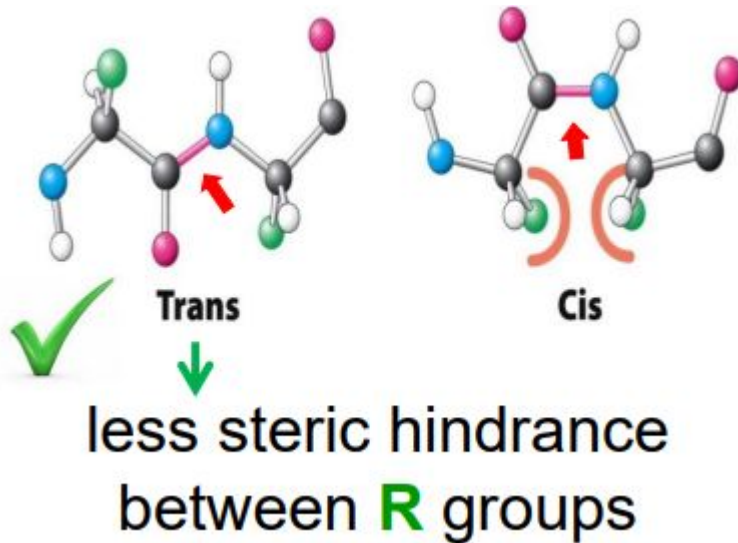
# SECONDARY STRUCTURE OF PROTEIN

- conformation of a region of a peptide → **Secondary** (e.g. alpha helix)
- conformation of a peptide as one entity → **Tertiary** (e.g. myoglobin)
- conformation of multiple peptide → **Quaternary** (e.g. hemoglobin)



# PEPTIDE ANGLE

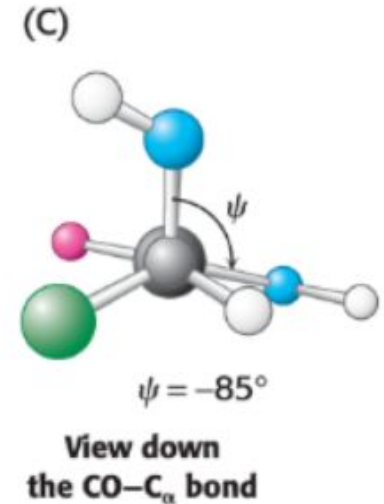
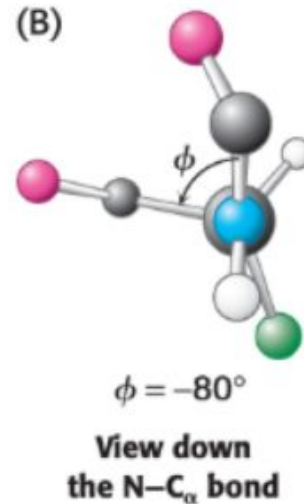
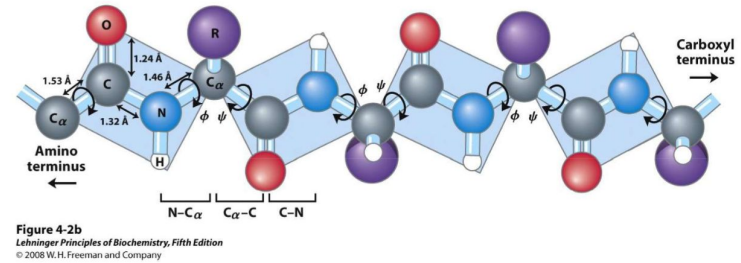
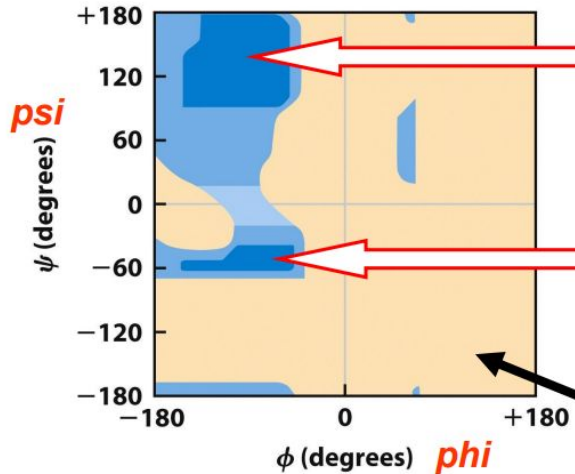
- peptide bond (HN-CO) = semi-planar, no rotation
- Usually, peptide bond adapt trans conformation
- Cis is seldomly allowed (except when Proline involved)





# RAMACHANDRAN PLOT

- Phi  $\phi$  :  $\text{C}(\text{O})-\text{N}-\text{C}(\alpha)-\text{C}(\text{O})$  bonds
- Psi  $\psi$  :  $\text{N}-\text{C}(\alpha)-\text{C}(\text{O})-\text{N}$  bonds
- clockwise: positive
- anticlockwise: negative



reverse  $\rightarrow$  negative  
when clockwise

# ALPHA HELIX

- bury peptide backbone
- side chains pointing outside
- Hydrogen bond between CO and NH among backbone
- Interaction between n and n+3/4 residue (facing same direction)
- compact

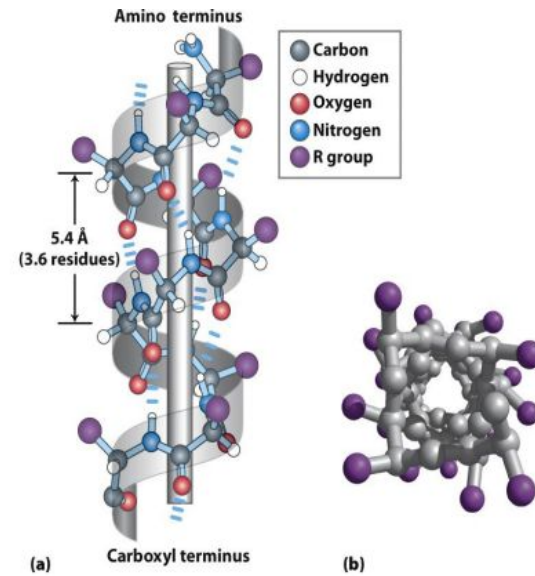
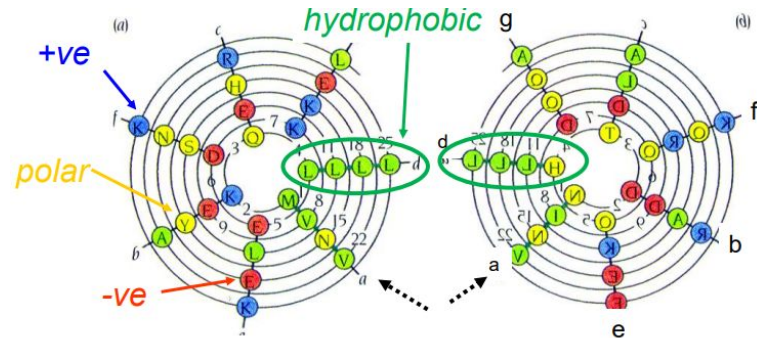


Figure 4-4  
Lehninger Principles of Biochemistry, Fifth Edition  
© 2008 W. H. Freeman and Company

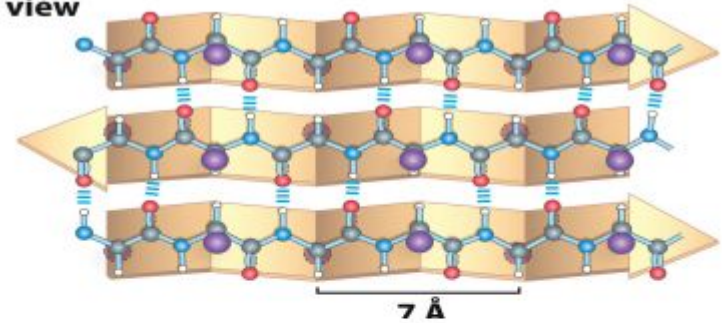


# BETA STRAND

- Parallel or antiparallel(anti more stable)
- side chain pointing alternating direction
- more extended
- zig-zag, not a flat strand

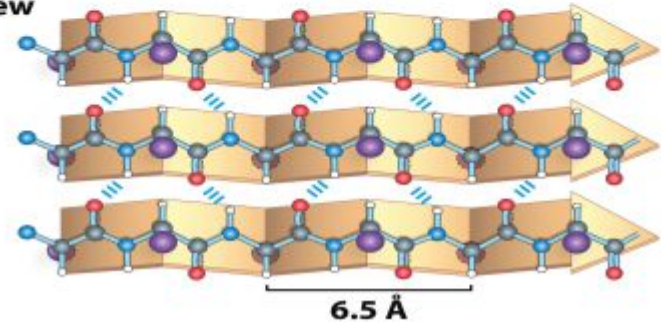
**Antiparallel  $\beta$  sheet**

**Top view**



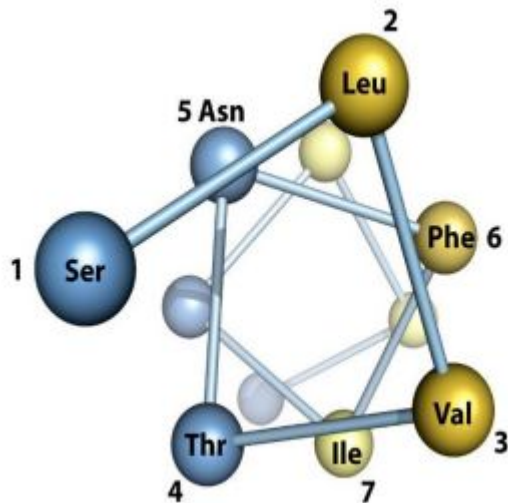
**Parallel  $\beta$  sheet**

**Top view**



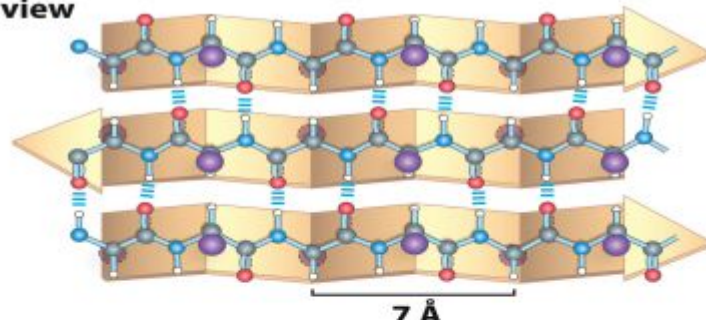
# DEDUCE SECONDARY STRUCTURE FROM SEQUENCE

- NOT GUARANTEED!!! (might have exception)
- Consider globular protein only (membrane and fibrous excluded) → **amphipathic** secondary structure
- Consider distribution of amino acids, not singular AA



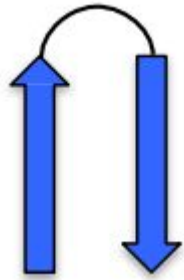
**Antiparallel  $\beta$  sheet**

**Top view**



# TERTIARY AND QUATERNARY STRUCTURE

- Secondary structures groups together -> forming motif



**beta  
hairpin**



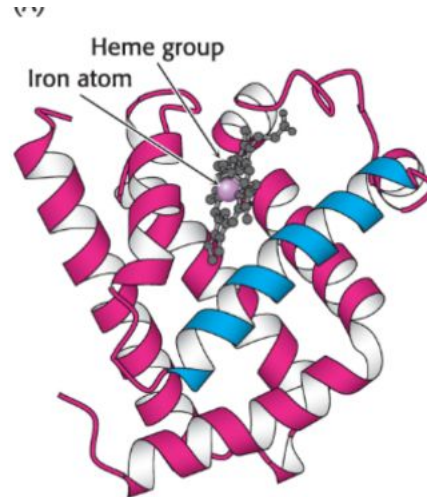
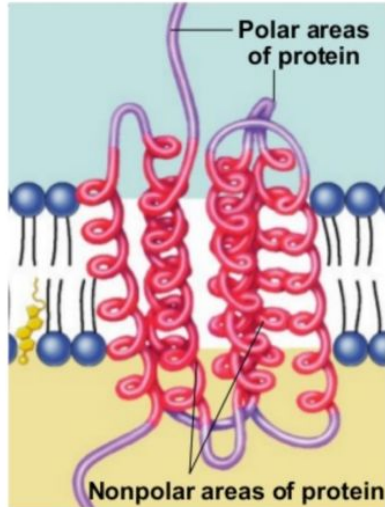
**beta-alpha-beta  
motif**

- One protein can have more than 1 motifs

# TERTIARY AND QUATERNARY STRUCTURE

- covalent (S-S) and non-covalent interaction
  - disulphide bond, hydrophobic interaction, charge interaction, Hydrogen bond between side chains
- Mostly related to side chains, not backbone
- May involved other groups (heme, FAD, lipoate ...)

membrane  
protein  
(contact  
with  
hydrophobic  
fatty acid)



globular protein  
(contact with  
hydrophilic  
environment)

# QUIZ 1

Which of the following amino acids would you expect to see in a protein present in the transmembrane region?

- a) Arginine
- b) Leucine
- c) Glutamine
- d) Histidine

# QUIZ 1

Which of the following amino acids would you expect to see in a protein present in the transmembrane region?

- a) Arginine (positively charged polar amino acid)
- b) Leucine (non-polar amino acid)
- c) Glutamine (neutral and polar amino acid)
- d) Histidine (positively charged polar amino acid)



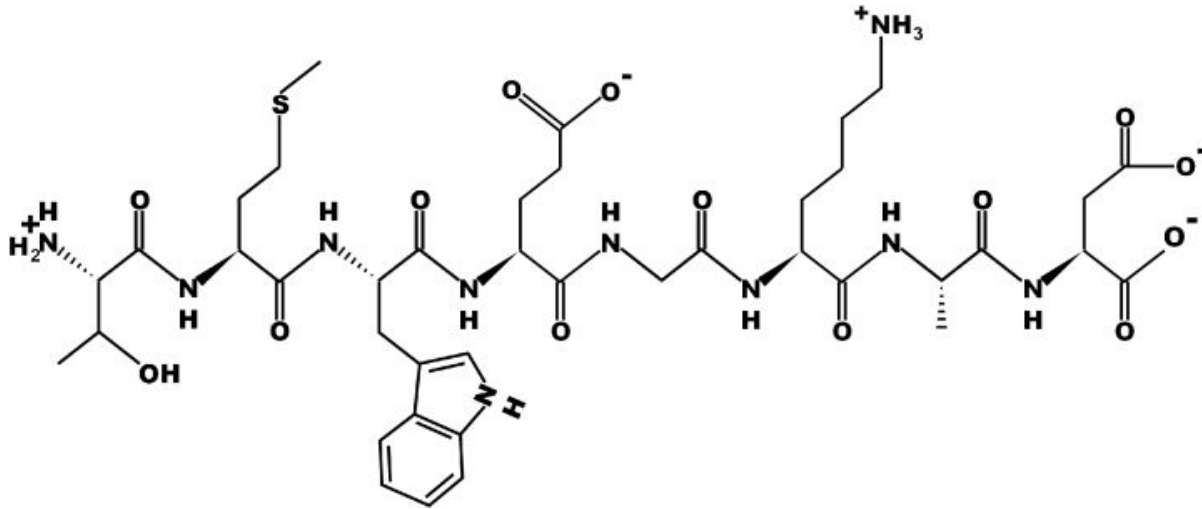
# PEPTIDE EXAMPLE

What is the charge of the following peptide chain at physiological pH (pH = 7.4)?

TMWEGKAD

# PEPTIDE EXAMPLE

What is the charge of the following peptide chain at physiological pH (pH = 7.4)?



# AMINO ACID SEQUENCE

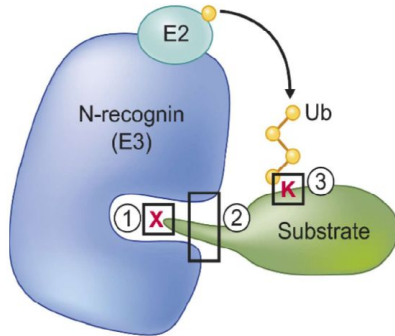
Protein sequence presented using the **abbreviations** of the 20 amino acids

- It is directional!
  - (N-end)TMWEGKAD(C-end) is different from (N-end)DAKGEWMT(C-end)
  - One example about “end-specificity”: N-end rule
- We can BLAST the sequence of a protein to find other proteins with similar primary structure (and very often, they will be its homologous proteins)

# DIRECTIONAL: N-END RULE (NO NEED TO KNOW DETAILS; IT'S IN CMBI4001)

## *The N-end Rule: in-vivo half-life of proteins*

- The **N-terminus of a protein** can undergo a diverse posttranslational modifications
  - create **degradation signal (N-degron)** that mediate protein destruction via **the N-end rule pathway** of ubiquitin-mediated proteolysis.
- **N-recognin:** The E3 Ub ligases that recognize N-degron



Three key features that determine an **N-degron**:

1. a primary N-terminal **destabilizing amino acid ('X')**;
2. an unstructured N-terminal region ensuring the N-residue is exposed and accessible;
3. an appropriately positioned downstream **lysine(s) (K)** to act as a receptor site for ubiquitin conjugation.

**Take-away message: N terminus and C terminus has their own function respectively.**

So (N-end) TMWEGKAD  
(C-end) is definitely different from  
(N-end) DAKGEWMT  
(C-end).

# BLASTP

[https://blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastp&PAGE\\_TYPE=BasicSearch&LINK\\_LOC=blasthome](https://blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastp&PAGE_TYPE=BasicSearch&LINK_LOC=blasthome)

to blast for protein sequence!

BLAST = basic local alignment search tool

Do you know what protein “mkalivlglv llsvtvqgkv fercelartl  
krlgmdgyrg islanwmcla kwesgynttra tnynagdrst dygifqinsr ywcndgktpg  
avnachlscs allqdniada vacakrvvrd pqgirawvaw rnrcqnrdvr qyvqgcgv”  
is? What proteins have similar sequence to it?

Blast it!

# BLASTP

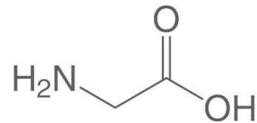
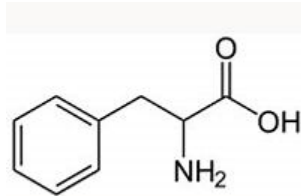
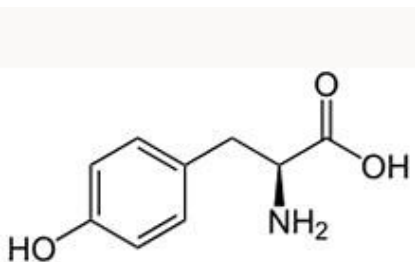
Sequences producing significant alignments									
Download ▾ Select columns ▾ Show 100 ▾ ?									
<input checked="" type="checkbox"/> select all 100 sequences selected         GenPept Graphics Distance tree of results Multiple alignment MSA Viewer									
	Description ▾	Scientific Name ▾	Max Score ▾	Total Score ▾	Query Cover ▾	E value ▾	Per. Ident ▾	Acc. Len ▾	Accession
<input checked="" type="checkbox"/>	lysozyme_C precursor [Homo sapiens]	<a href="#">Homo sapiens</a>	303	303	100%	5e-104	100.00%	148	<a href="#">NP_000230.1</a>
<input checked="" type="checkbox"/>	lysozyme_precursor (EC 3.2.1.17) [Homo sapiens]	<a href="#">Homo sapiens</a>	303	303	100%	1e-103	99.32%	148	<a href="#">AAA36188.1</a>
<input checked="" type="checkbox"/>	LYZ [synthetic construct]	<a href="#">synthetic construct</a>	302	302	100%	3e-103	99.32%	148	<a href="#">AKI70658.1</a>
<input checked="" type="checkbox"/>	lysozyme_C precursor [Gorilla gorilla]	<a href="#">Gorilla gorilla</a>	301	301	100%	3e-103	99.32%	148	<a href="#">NP_001266591.1</a>
<input checked="" type="checkbox"/>	lysozyme [synthetic construct]	<a href="#">synthetic construct</a>	301	301	100%	4e-103	99.32%	148	<a href="#">BAG73364.1</a>
<input checked="" type="checkbox"/>	LYZ [synthetic construct]	<a href="#">synthetic construct</a>	301	301	100%	4e-103	99.32%	148	<a href="#">AKI70659.1</a>
<input checked="" type="checkbox"/>	lysozyme_C [Pongo abelii]	<a href="#">Pongo abelii</a>	300	300	100%	2e-102	97.97%	148	<a href="#">XP_002823550.1</a>
<input checked="" type="checkbox"/>	LYZ [synthetic construct]	<a href="#">synthetic construct</a>	300	300	100%	2e-102	99.32%	148	<a href="#">AKI70661.1</a>
<input checked="" type="checkbox"/>	LYZ [synthetic construct]	<a href="#">synthetic construct</a>	300	300	100%	2e-102	99.32%	148	<a href="#">AKI70660.1</a>
<input checked="" type="checkbox"/>	lysozyme_C [Nomascus leucogenys]	<a href="#">Nomascus leuco...</a>	298	298	100%	6e-102	96.62%	148	<a href="#">XP_003259554.1</a>
<input checked="" type="checkbox"/>	lysozyme_precursor [Homo sapiens]	<a href="#">Homo sapiens</a>	297	297	100%	2e-101	96.62%	148	<a href="#">AAC63078.1</a>
<input checked="" type="checkbox"/>	RecName: Full=Lysozyme C; AltName: Full=1.4-beta-N-acetylmuramidase C; Flags: Precursor [Hylobates lar]	<a href="#">Hylobates lar</a>	295	295	100%	2e-100	95.27%	148	<a href="#">P79180.1</a>
<input checked="" type="checkbox"/>	lysozyme_C [Chlorocebus sabaeus]	<a href="#">Chlorocebus sa...</a>	283	283	100%	6e-96	89.86%	148	<a href="#">XP_008002172.1</a>
<input checked="" type="checkbox"/>	RecName: Full=Lysozyme C; AltName: Full=1.4-beta-N-acetylmuramidase C; Flags: Precursor [Nasalis larva]	<a href="#">Nasalis larvatus</a>	280	280	100%	1e-94	87.84%	148	<a href="#">P79811.1</a>
<input checked="" type="checkbox"/>	lysozyme_C precursor [Macaca mulatta]	<a href="#">Macaca mulatta</a>	280	280	100%	2e-94	88.51%	148	<a href="#">NP_001095203.1</a>
<input checked="" type="checkbox"/>	RecName: Full=Lysozyme C; AltName: Full=1.4-beta-N-acetylmuramidase C; Flags: Precursor [Miopithecus tala...	<a href="#">Miopithecus tala...</a>	279	279	100%	2e-94	88.51%	148	<a href="#">P79806.1</a>
<input checked="" type="checkbox"/>	lysozyme_C precursor [Papio anubis]	<a href="#">Papio anubis</a>	279	279	100%	2e-94	88.51%	148	<a href="#">NP_001106112.1</a>
<input checked="" type="checkbox"/>	RecName: Full=Lysozyme C; AltName: Full=1.4-beta-N-acetylmuramidase C; Flags: Precursor [Pygathrix nema...	<a href="#">Pygathrix nema...</a>	278	278	100%	4e-94	86.49%	148	<a href="#">P79847.1</a>
<input checked="" type="checkbox"/>	RecName: Full=Lysozyme C; AltName: Full=1.4-beta-N-acetylmuramidase C; Flags: Precursor [Semnopithecus ...]	<a href="#">Semnopithecus ...</a>	278	278	100%	1e-93	86.49%	148	<a href="#">P67977.1</a>

Answer: That sequence is **human's lysozyme C precursor!** You can also see that in the result list, some homologous proteins have good match to it.

# HOMOLOGOUS PROTEINS SHARE SIMILAR SEQUENCE

## Residues:

- **invariant** (totally conserved): the amino acid residue is identical for **ALL** species
- **variant**: the amino acid residue is NOT totally identical
  - **conservative substitutions**: residues are replaced by amino acids of **similar chemical properties**

[illegible]

**FYI:** The BLOSUM62 matrix is derived from comparing amino acid substitutions among proteins that are no more than 62% similar – it's derived from empirical data

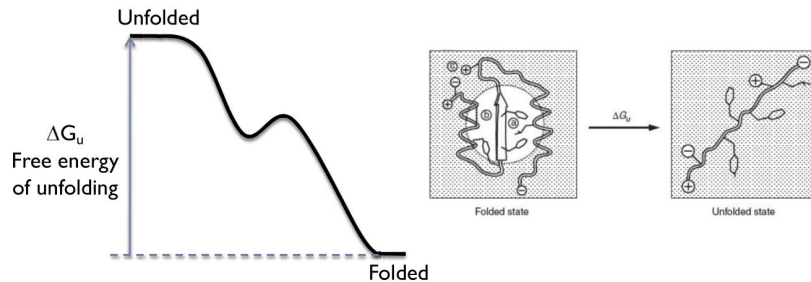
Henikoff, S. & Henikoff, J.G. (1992)  
Proc. Natl. Acad. Sci. USA  
89:10915-10919.

# WHAT DETERMINES PROTEIN TERTIARY STRUCTURE?

Protein's **primary structure** has enough information dictating its folding (you will learn a lot about this in CMBI4002)

## Thermodynamics hypothesis

- ▶ Anfinsen (1972) "The three-dimensional structure of a native protein in its normal physiological milieu is the one in which the **Gibbs free energy of the whole system is lowest**; that is, the native conformation is determined by the totality of inter-atomic interactions and hence by the amino acid sequence"
- ▶ The tendency to reach this free energy minimum drives proteins fold.



Stabilizing factors:

hydrophobic effect

hydrogen bond

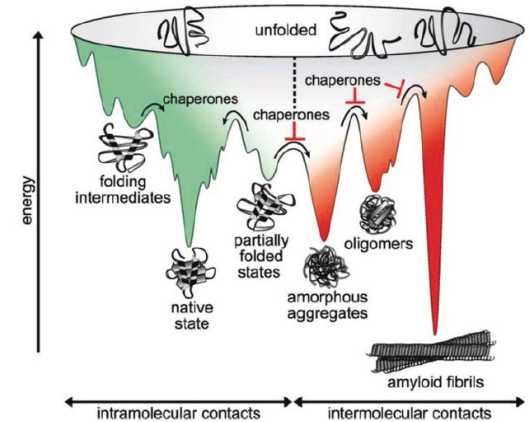
electrostatic interaction

(these weak interactions make protein fold properly;  
**urea can disrupt some of these interactions**)



# SOME FEATURES OF PROTEIN FOLDING

- **stepwise** process, NOT random trial-and-error
- **intramolecular** interactions drive proteins towards its functional native state (**thermodynamically favourable**)
- partially folded intermediates may either: 1) turn to aggregates (often disease-causing) due to **intermolecular** interactions, or 2) assisted by chaperones, fold back to the native conformation



# QUIZ 2

Which statement best explains why antiparallel beta strands are more stable?

- a) Antiparallel beta strands are highly packed
- b) Parallel beta strands form fewer hydrogen bonds between adjacent strands
- c) Antiparallel beta strands form stronger hydrogen bonds between adjacent strands
- d) Parallel beta strands experience greater steric hindrance due to the orientation of R groups.

# QUIZ 2

Which statement best explains why antiparallel beta strands are more stable?

- a) Antiparallel beta strands are highly packed
- b) Parallel beta strands form fewer hydrogen bonds between adjacent strands
- c) Antiparallel beta strands form stronger hydrogen bonds between adjacent strands (because the C=O and NH group of peptide bonds in adjacent strands are closer)
- d) Parallel beta strands experience greater steric hindrance due to the orientation of R groups. (R groups extend from sheet and point on alternatively on opposite sides)